

Nutritional value and antioxidant activity of ora-pro-nobis, tamarillo and green banana biomasses Valor nutricional e atividade antioxidante de ora-pro-nóbis, tamarillo e

biomassas de bananas verdes

Article Info:

Article history: Received 2024-10-10 / Accepted 2024-12-08 / Available online 2024-12-11 doi: 10.18540/jcecvl10iss8pp20740



Renata de Souza Ferreira ORCID: https://orcid.org/0000-0001-5622-2540 Universidade Federal de Viçosa and IF Sudeste MG - campus Barbacena, Brazil E-mail: renata.s.ferreira@ufv.br **Jucenir dos Santos** ORCID: https://orcid.org/0000-0001-6911-4354 Universidade Federal de Viçosa, Brazil E-mail: jucenir.santos@ufv.br Lívva Alves de Oliveira Universidade Federal de Viçosa, Brazil ORCID: https://orcid.org/0000-0003-0206-633X Universidade Federal de Viçosa, Brazil E-mail: lyvia.oliveira@ufv.br **Ceres Mattos Della Lucia** ORCID: https://orcid.org/0000-0002-6731-5694 Universidade Federal de Viçosa, Brazil E-mail: cmdellalucia@ufv.br Márcia Cristina Teixeira Ribeiro Vidigal ORCID: https://orcid.org/0000-0002-8065-0753 Universidade Federal de Vicosa, Brazil E-mail: marcia.vidigal@ufv.br

Abstract

The consumption of unconventional food plants (UFP) can be a strategy to promote food and nutritional security, for which it is necessary to know their nutritional composition. The objective of this study was to analyze the nutritional value and antioxidant activity of ora-pro-nobis, tamarillo and bananas biomass (*prata* and *nanica*). Using standard methodologies (AOAC, 2005), the following compounds were analyzed in triplicate: protein, lipids, carbohydrate, calories, moisture, total fiber content, calcium, iron, vitamin C (by titrimetry), carotenoids (by High Performance Liquid Chromatography), phenolic compounds (by Folin-Ciocalteau) and antioxidant activities, by DPPH (2,2-diphenyl-1-picrylhydrazyl) and ABTS (2,2'-azino-bis 3-ethylbenzothiacholine-6-sulfonic acid). Ora-pro-nobis stands out as a source of calcium, iron, carotenoids and protein, is low in fat and has good antioxidant activity. Tamarillo showed high antioxidant activity, is a protein source and low in fat. The two banana biomasses had similar results in most nutrients, but the "nanica" banana with more protein and the "prata" with greater antioxidant activity. The unconventional food plants analyzed are potential bioactive foods, because they have phenolic compounds and high antioxidant activity. Thus, UFP have the potential to promote a more diverse and nutritious diet for consumers in general.

Keywords: *Pereskia aculeata. Solanum betaceum. Musa paradisíaca.* Unconventional food plants. Bioactive compounds.

Resumo

O consumo de plantas alimentícias não convencionais (PANC) pode ser uma estratégia para promover a segurança alimentar e nutricional, para a qual é necessário conhecer sua composição nutricional. O objetivo deste estudo foi analisar o valor nutricional e a atividade antioxidante de orapro-nóbis, tamarillo e das biomassas de bananas (prata e nanica). Utilizando metodologias padrão (AOAC, 2005), os seguintes compostos foram analisados em triplicata: proteína, lipídios, carboidrato, calorias, umidade, teor total de fibras, cálcio, ferro, vitamina C (por titulação), os carotenoides (por Cromatografia Líquida de Alta Eficiência), os compostos fenólicos (por Folin-Ciocalteau) e as atividades antioxidantes, por DPPH (2,2-difenil-1-picril-hidrazil) e ABTS (ácido 2,2'-azino-bis 3-etilbenzotiacolina-6-sulfônico). Ora-pro-nóbis se destaca como fonte de cálcio, ferro, carotenoides e proteína, é pobre em gordura e tem boa atividade antioxidante. O tamarillo apresentou alta atividade antioxidante, é fonte de proteína e pobre em gordura. As duas biomassas de banana tiveram resultados tiveram resultados similares na maioria dos nutriente, porém a banana nanica com mais proteína e a prata com maior atividade antioxidante. As plantas alimentícias não convencionais analisadas são alimentos bioativos em potencial, porque apresentam compostos fenólicos e alta atividade antioxidante. Desta forma, as PANCs têm potencial para promover uma dieta mais diversificada e nutritiva para os consumidores em geral.

Palavras-chave: Pereskia aculeata. Solanum betaceum. Musa paradisíaca. Planta alimentícia não convencional. Compostos bioativos.

1. Introduction

Fruits and vegetables should be included in daily diets as they promote health and help prevent various diseases (Liu, 2013). However, in Brazil, there are more than 3,000 species of Unconventional Food Plants (UFP) with food potential that remain unknown or are not consumed by most of the population. Some examples include ora-pro-nobis (*Pereskia aculeata*), tamarillo (*Solanum betaceum*), and unconventional parts of common or conventional plants, such as green banana fruit (*Musa paradisíaca*) (Kinupp & Lorenzi, 2014).

Pereskia aculeata is popularly known as ora-pro-nobis, *lobrobô*, Barbados gooseberry, among other names. Its leaves, flowers, and fruits are edible (Kinupp & Lorenzi, 2014) and have attracted interest from the food and pharmaceutical industries due to its bioactive compounds and mucilage (Ferreira *et al.*, 2024; Porto *et al.*, 2021).

Solanum betaceum is popularly known as tamarillo, tree tomato, Indian tomato, or French tomato. It has edible fruit (Kinupp & Lorenzi, 2014) and contains high levels of bioactive compounds, fiber, and protein, as well as antioxidant and anti-hyperglycemic activities, giving it functional value beyond its nutritional value (Orqueda *et al.*, 2021).

The banana (*Musa paradisíaca*) is a conventional fruit; however, other parts of this plant are generally not consumed (unconventional), such as the green fruit (Kinupp & Lorenzi, 2014). Green bananas are a source of minerals, vitamins, and resistant starch, which is beneficial against intestinal cell cancer, heart disease, and celiac disease (Feitosa *et al.*, 2023). However, studies on quantification of phenolic compounds (Riquette *et al.*, 2019; Sena *et al.*, 2020) are still scarce, especially with the species "nanica" (Riquette *et al.*, 2019) and "prata" (Silva *et al.*, 2016). None of this analyzed antioxidant activity.

Additionally, studies that establish daily consumption recommendations for these plants based on their composition are scarce, as are studies analyzing the antioxidant activity of these plants and its correlation with bioactive compounds such as phenolic compounds, carotenoids, and vitamin C.

The consumption of UFP can be a strategy for promoting food and nutritional security, as it encourages a more diverse diet and helps reduce nutritional deficiencies (Jacob *et al.*, 2020; MAPA, 2010; Barreira *et al.*, 2015). To achieve this, it is necessary to understand their nutritional information and quantify their compounds. In this context, this study aimed to analyze the selected UFP (ora-pro-nobis, tamarillo, and green banana biomass) regarding their levels of proteins, lipids, carbohydrates, moisture, total fiber, calcium, iron, vitamin C, carotenoids, phenolic compounds, and antioxidant activity.

2. Methodology

2.1 Chemical products and reagentes

All reagents are of analytical grade: 2,2-diphenyl-1-picrylhydrazyl radical; 2,2'-azino-bis radical (3-ethylbenzothiacholine-6-sulfonic acid); methanol; gallic acid; ethanol; Trolox; ethyl acetate; acetonitrile; metaphosphoric acid; 2,6-dichlorophenol indophenol; standard ascorbic acid; nitric acid; perchloric acid; acetic acid; trichloroacetic acid; petroleum ether; sulfuric acid; potassium sulfate; copper sulfate; sodium hydroxide; boric acid; Tashiro indicator; hydrochloric acid.

2.2 Equipment, instruments and glassware

Analytical balance (precision of 0.0001 g), heating chamber, muffle furnace, water bath, electric plate, nitrogen distiller (Solab SL-74®), Soxhlet apparatus (Marcon®), rotary evaporator, blender, mechanical shake, spectrometer (ICP-OES; Perkin Elmer Model Optima 8300 DV), high-performance liquid chromatography (HPLC) system, refrigerator, freezer. Desiccator, porcelain capsule, Kjeldahl flask, distillation flask, Erlenmeyer flask, burette, graduated and volumetric pipettes, flat-bottomed flask, glass chromatographic column, kitassatos, glass rod, glass funnel, volumetric flasks, Büchner funnel, measuring cylinder, beakers. Tweezers, metal spatula, tissue paper and cotton.

2. 3 Plant material

The plants were obtained from family farmers in the Zona da Mata region of Minas Gerais, Brazil. ora-pro-nobis and tamarillo (Figure 1) were harvested in the municipality of Brás Pires (20°50'52"S43°14'39"W) and *prata* and *nanica* bananas were obtained in Viçosa – MG (20°47'40"S42°56'11"W). The samples were transported in a polyethylene box protected from light. Then, it was washed in running water, sanitized for 15 min in a 100 ppm chlorinated solution and rinsed in filtered water. Excess moisture was removed with paper towels.



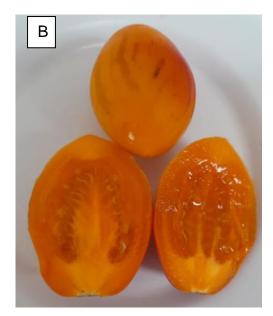


Figure 1 - Ora-pro-nobis (A) and tamarillo (B)

The biomass was prepared by cooking the whole green bananas for 8 min under pressure, using the proportion of water: green banana 1:1 (v: w). The boiling water was added and after 8 min of pressure, the pressure was released naturally. While still hot, the banana was peeled and blended until homogenized. In this way, the biomass was composed of cooked green banana pulp. (Figure 2).

The samples were stored protected from light, under refrigeration (5°C to 8 °C), for up to 7 days or in the freezer (-12 °C to -18 °C), for up to 90 days. Analyzes were performed in triplicate.



Figure 2 - Green banana biomass processing.

Note: A – Washing; B – Sanitize; C – Put in the pan: green banana and water (1:1); D – Cooking under pressure; E – Drain cooking water; F – Peel; G – Process until homogenized; H – Store.

2. 4 Nutritional composition

The nutritional composition of the plants was determined following the methods recommended by the Association of Official Analytical Chemicals (AOAC, 2005).

Moisture was determined by direct drying in an oven at 105 °C until constant final weight (variation less than 0.01 g). Ash or fixed mineral residue was obtained by incineration in a muffle furnace at 550 °C until constant final weight (variation less than 0.01 g).

Protein was determined by the modified Kjeldahl method. The digester solution contained sulfuric acid, potassium sulfate and copper sulfate. For distillation, 40 % (v/v) sodium hydroxide, 4 % (v/v) boric acid and Tashiro indicator were used in a nitrogen distiller (Solab SL-74 \mathbb{B}). The titration was carried out with 0.01 N hydrochloric acid (f(HCl) = 0.994) and Tashiro indicator. The nitrogen to protein conversion factor adopted was 6.25.

The lipids were extracted, continuously for 6 h, in a Soxhlet apparatus (Marcon®), using petroleum ether as the solvent. The crude fiber was determined by acid digestion (with acetic, nitric and trichloroacetic acids), followed by incineration (at 550 °C) of the sample until only crude fiber.

The carbohydrate content was calculated by difference using the equation: [100 - (% moisture + % lipid + % protein + % total fiber + % ash)] (Oliveira *et al.*, 2019). The caloric value was calculated based on the contents of the protein, lipid, and carbohydrate fractions, using specific coefficients that consider the heat of combustion 4.0; 9.0 and 4.0 kcal, respectively (Dutra de Oliveira; Marchini, 1998).

2. 5 Determination of calcium and iron

For calcium and iron analysis, the samples were dried in forced air circulation ovens for 72 h at a temperature between 68 and 72 °C, and subsequently homogenized by grinding. Digestion was carried out with nitric acid and perchloric acid, on a preheated plate at 80 °C until reaching 200 °C (Sarruge; Haag, 1974). The analysis was performed on an inductively coupled plasma optical emission spectrometer (ICP-OES; Perkin Elmer Model Optima 8300 DV). For quantification, analytical curves were constructed using mineral standards. The results were converted into mineral concentrations, considering the dilutions and their possible difference about in relation to the blank.

2. 6 Determination of C vitamin

Vitamin C was determined using the Tillmans method. The sample of known weight was placed in a beaker with the metaphosphoric acid solution and titrated with the 2,6-dichlorophenol indophenol solution, compared with the standard ascorbic acid solution. The adaptation was carried out with ora-pro-nobis. Due to its solid consistency in its natural state, it was previously ground, diluted in water (1:10, weight), and filtered for subsequent analysis. Vitamin C content was calculated using the Equation 1:

$$Vitamin \ C \ content = \frac{V * F}{A} * 100 \tag{1}$$

Where V is the titration volume, F is the Tillman factor (mass of vitamin C used in titration)/(volume of titrant used in standard titration), and A is the sample volume after titrant addition.

2. 7 Determination of carotenoids

For carotenoid analysis, the samples and extracts were protected from both sunlight and artificial light with the use of amber glass bottles, aluminum foil and blackout curtains. To extract the carotenoids, five grams of sample were added 20 mL of acetone and homogenized in a microshredder for 3 min and vacuum filtered. This procedure was repeated 3 times. Then, the filtrate was added to 50 mL of cooled petroleum ether and washed with distilled water to remove acetone. To the carotenoid extract in petroleum ether kept in the funnel, anhydrous sodium sulfate was added to remove residual water. Finally, the extract was concentrated to 10 mL, using a rotary evaporator (Rodriguez-Amaya, 2001). The analysis was carried out using a high-performance liquid chromatography (HPLC) system, at a wavelength of 450 nm (Pinheiro-Sant'Ana et al., 1998). The liquid chromatography system used featured a Shimadzu® diode array detector and a Gemini reversed-phase analytical column (250 × 4 mm, 5 µm) preceded by an ODS safety pre-column (C18) $(4.0 \times 3.0 \text{ mm})$, both from Phenomenex® (Torrance, CA, USA). The mobile phase was composed of methanol, ethyl acetate and acetonitrile, (80:10:10). The identification of carotenoids was made by comparing their UV spectra and retention times with reference standards. Quantification was carried out using a standard curve established from the same standards. The result was expressed in mg of carotenoid/100 g.

2. 8 Extraction and determination of the phenolic compounds

Total phenolic content was determined using the Folin-Ciocalteu method. Extraction of the compounds was carried out with 1 g of sample in a methanol solution: water 60:40 (v/v), under agitation for 30 min and subsequent centrifugation at 3500 rpm for 5 min (Bloor, 2001). Phenolic compounds were determined by reading on a spectrophotometer, at 765 nm absorbance, based on an analytical curve of gallic acid (0.0025 - 0.03 g/L), previously carried out. The results were expressed as mg of gallic acid equivalent per g of sample (mg GAE/ g) (Singleton *et al.*, 1999).

2. 9 Antioxidant activity

Antioxidant activity was determined by 2 methods: DPPH, with 2,2-diphenyl-1picrylhydrazyl radical and ABTS, with 2,2'-azino-bis radical (3-ethylbenzothiacholine-6-sulfonic acid). Extraction was carried out according to Bloor (2001).

Using the DPPH method, methanolic DPPH solution was added to the extracts, and the extracts, blank and control were read in a spectrophotometer, at 517 nm absorbance, based on a previously performed Trolox analytical curve. The percentage of radical inhibition was calculated using the Equation 2:

% radical inhibition =100 -
$$\frac{(A_f/A_0)}{A_c}$$
 * 100 (2)

Where " A_0 " is the initial absorbance, " A_f " is the final absorbance, "Ac" is the control absorbance (Blois, 1958).

In the ABTS method, the extraction of compounds was carried out in a 60% methanol solution. The extract was evaporated at 44°C for about 15 min, until the methanol dried. Next, 95° PA ethanol was added, the same solvent as the standard Trolox solution. The antioxidant activity was determined by reading on a spectrophotometer, at 734 nm absorbance, based on a previously performed Trolox analytical curve (100 – 2000 μ mol/L) (Boroski *et al.*, 2015; Brasil, 2007). ABTS and DPPH results were expressed in mg/g.

2. 10 Analysis of results

This study was conducted in a completely randomized design with three replicates. Descriptive statistics (mean and standard deviation) were used to present the results. To compare the two banana species, the results of the analyses were compared by analysis of variance, using the One-Way ANOVA test ($p \le 0.05$). Data on antioxidant activity and bioactive compounds, such as phenolics and carotenoids, were correlated using Pearson's test. Statistical analyses were performed using R software.

The UFP were classified as "source," "good source," and "excellent source" (Philippi, 2008) when they had the potential to provide 5 to 10% of the dietary reference intake (DRI, according to the Institute of Medicine, 2020) in a serving; 10 to 20% of the potential DRI; and more than 20% of the DRI, respectively, as indicated by Philippi (2008), considering the portion of vegetables equivalent to 30 kcal and fruits equivalent to 70 kcal, as proposed by the dietary guidelines for the Brazilian population (Brasil, 2008).

3. Results and Discussion

3.1 Nutritional composition and antioxidant activity of ora-pro-nobis

The recommended daily portion for consumption of ora-pro-nobis was 79 g (Table 1), equivalent to 30 kcal (Brasil, 2008). This vegetable can be considered a source of protein, a good source of vitamin C, and an excellent source of calcium, iron, and carotenoids (5.46%, 18.93%, 100%, 45.25%, and 74.45% DRI/serving, respectively) while being low in lipids (0.34%), according to the daily recommended intake (DRI) proposed by the Institute of Medicine (2020). Additionally, it contains 37.89 kcal/100g, 87.45% moisture, 2.17% ash, 1.3% fiber, and 5.24% carbohydrates (Table 1). Ora-pro-nobis showed a higher protein content (3.46 \pm 0.06 g/100g) compared to other studies found in the literature, which reported values ranging from 1.27 to 2.8 g/100g (Barreira *et al.*, 2021 and Oliveira *et al.*, 2019, respectively). Ora-pro-nobis is considered a good source of plant protein, with a composition similar to that of legumes like beans (Silveira *et al.*, 2020), containing all essential amino acids, an abundant supply of tryptophan and lysine, but limited in methionine and cysteine (Santos *et al.*, 2022; Silveira *et al.*, 2020; Takeiti *et al.*, 2009; Zem *et al.*, 2017). Using different protein sources can improve nutritional quality. In this case, to complement amino acids, it is suggested to consume it alongside cereals, as they are rich in the limiting amino acids found in ora-pro-nobis (Silveira *et al.*, 2020).

The content of phenolic compounds and antioxidant activity by ABTS of ora-pro-nobis (Table 1) was also higher than that found in the literature (Table 2). Ora-pro-nobis showed a low lipid content (Table 1), lower than that reported in the literature (Table 2).

The crude fiber and vitamin C levels found in ora-pro-nobis were lower than those available in the literature. It is worth noting that the methodology of this study differs from some other studies that used HPLC for vitamin C and determined dietary fiber rather than crude fiber. The methodologies for crude fiber and vitamin C by Tillmans (AOAC, 2005) are validated and have the advantages of being less costly, faster, and more practical.

The ora-pro-nobis leaves presented a caloric value (Table 1) similar to other leafy vegetables such as broccoli (34 kcal/100 g), cauliflower (25 kcal/100 g), and cabbage (31 kcal/100 g) (Favela-González *et al.*, 2020). However, its fiber content (Table 1) is higher than that of broccoli (2.6 g/100 g), cauliflower (2.0 g/100 g), and cabbage (2.2 g/100 g) (Favela-González *et al.*, 2020). Regarding

protein content (Table 1), ora-pro-nobis leaves have higher values compared to leafy vegetables like cabbage. (1,28 g/100 g) (Brito *et al.*, 2020).

Parameters	Ora-pro-nobis	Ora-pro-nobis	%DRI*
	(in 100 g)	(portion: 79 g)	
Calories (kcal)	37.89±1.17	29.94	-
Moisture (% m/m)	87.45±0.07	69.08	-
Ash (% m/m)	2.17±0.14	1.71	-
Protein (% m/m)	3.46±0.06	2.73	5.46
Lipids (% m/m)	0.34±0.09	0.27	-
Crude Fiber (% m/m)	1.3±0.1	1.03	4.12
Carbohydrate (% m/m)	5.24±0.19	4.14	3.18
Calcium (mg/ 100g)	3384 ± 11.13	2673	> 100
Iron (mg/ 100g)	8.02 ± 0.18	6.34	45.25
Vitamin C (mg/ 100g)	23.97 ± 3.75	18.94	18.93
Carotenoids (mg/ 100g)	62.837±5.871	49.64	74.45
Total phenolic content (mg GAE/ 100g)	180.76±6.63	-	-
Antioxidant activity, DPPH (µmol trolox/g of	6875±187; 87.19±2.37	-	-
sample; % AA; mg/g)	%;1721±47		
Antioxidant activity ABTS (µmol trolox/g of	57.49±3.60;	-	-
sample; % AA; mg/g)	64.74±4.36%;14.39±0.9		

Table 1 - Nutritional composition, phenolic compounds, and antioxidant activity of ora-pronobis leaves.

Analysis on a wet basis, expressed as mean (of 3 repetitions) ± standard deviation. *% DRI: daily reference intake, based on a 30 kcal portion (79 g) and a daily diet of 2000 kcal (according to Institute of Medicina, 2020)

Table 2 - Nutritional composition, phenolic compounds, and antioxidant activity data of ora-
pro-nobis leaves available in the literature.

Parameters	Ora-pro-nobis (results found in other studies)
Calories (kcal)	22.62^{1} to 42.0^{2}
Moisture (%)	86.65 ¹ to 91.10±2.24 ³
Ash (%)	0.96 ± 0.01^3 to 2.90 ± 0.51^4
Protein (%)	1.27 ± 0.07^3 to 2.80 ± 0.11^4
Lipids (%)	0.40 ± 0.12^4 to 1.45 ± 0.01^3
Fiber (%)	3.73 ± 0.03^3
Total dietary fiber	3.881
Crude Fiber	-
Carbohydrate (%)	2.65^{1} to 6.70 ± 1.64^{4}
Calcium (mg/ 100g)	269.38^{1} to 6491.0 ± 132.5^{4}
Iron (mg/ 100g)	1.33^{1} to 24.1 ± 4.1^{4}
Vitamin C	192.21 ⁶ mg/ 100g
Vitamin C (spectrophotometric)	192.21 ⁶ mg/100g
Vitamin C by Tillmans (mg/ 100g)	-
Carotenoids	1190±230.0 to 2100±210.9 ⁵ mg/ 100g
Total phenolic content	7.86 ± 1.59^4 to 151.503 ± 334.5^7 mg GAE/100 g
Antioxidant activity, DPPH	$27 \pm <1^8$ mg of AAE/ g to 106.1 ± 3.9^9 (IC ₅₀ : µg/ml)
Antioxidant activity ABTS (µmol trolox/g of sample; % AA; mg/g)	$40.5 \pm 1 (IC_{50}: \mu g/ml)^{10}$

Studies with ora-pro-nobis: ¹-Botrel *et al.*, 2020; ²-Monteiro *et al.*, 2021; ³-Barreira *et al.*, 2021; ⁴-Oliveira *et al.*, 2019; ⁵-Agostini-Costa *et al.*, 2014; ⁶-Oliveira *et al.*, 2013; ⁷-Maciel *et al.*, 2021; ⁸-Cruz *et al.*, 2021; ⁹-Souza *et al.*, 2014; ¹⁰-Garcia *et al.*, 2019

3.2 Nutritional Composition and Antioxidant Activity of Tamarillo

The recommended daily portion for tamarillo consumption was 187.5 g (Table 3), equivalent to 70 kcal (Brazil, 2008). This fruit can be considered a source of protein, a good source of calcium, and an excellent source of fiber, vitamin C, and iron (7,33%; 17,89%; 21,92%; 39,54% e 37,39 % DRI/ portion, respectively) while being low in lipids (0.05%), according to the daily recommended intake (DRI) proposed by the Institute of Medicine (2020). Additionally, the tamarillo fruit contains: 37.33 Kcal/ 100g, 1.11% ash, and 2.9% carbohydrate (Table 3).

Parameters	Tamarillo (100g)	Tamarillo	%DRI*
		(portion: 187,5 g)	
Calories (kcal)	37.33±1.24	70.0	3.5%
Moisture (% m/m)	86.71±0.33	-	-
Ash (% m/m)	1.11±0.09	-	-
Protein (% m/m)	1.94±0.05	3.66	7.32%
Lipids (% m/m)	0.05±0.01	0.09	0.14%
Crude Fiber (% m/m)	2.9±0.1	5.43	21.72%
Carbohydrate (% m/m)	2.9±0.1	13.67	4.56%
Calcium (mg/ 100g)	94.66 ± 2.88	177.5	17.75%
Iron (mg/ 100g)	2.77 ± 0.06	5.19	37.07%
Vitamin C (mg/ 100g)	20.92 ± 0.11	39.23	39.23%
Carotenoids (mg/ 100g)	49.2±4.66	-	-
Antioxidant Activity, DPPH (mg/g; % AA)	191±29;	-	-
	20.81±1.72 %		
Antioxidant Activity ABTS (mg/g)	71.03±0.78	-	-

Table 3. Nutritional composition, phenolic compounds, and antioxidant activity of tamarillo fruit.

Analysis on a wet basis, expressed as mean (of 3 repetitions) \pm standard deviation. *%DRI: daily reference intake, based on a 70 kcal portion (163 g) and a daily diet of 2000 kcal (according to Institute Medicina, 2020).

For most parameters, the values found (Table 3) were intermediate compared to those mentioned in the literature (Table 4). For antioxidant activity by ABTS and ash content, higher values (Table 3) were obtained compared to those reported in the literature (Table 4).

In the present study, the characterization was performed on the whole fruit. However, Martin *et al.* (2021) observed differences in the chemical composition of different parts of the fruit, with seeds being richer in lipids, pectin, and phenolic compounds; the peel containing fiber, pectin, lipids, higher levels of phenolics, and terpenoids; and the pulp predominantly polysaccharides, fiber, pectin, and phenolic compounds.

Tamarillo has a lower caloric value (37.33 kcal/100 g, as shown in Table 3) compared to various fruits such as pineapple, "Prata" banana, "lima" orange, "Fuji" apple, "Formosa" papaya, "Palmer" mango, and "Italia" grape (48, 96, 46, 56, 45, 72, and 53 kcal/100 g, respectively) (TACO, 2011). The lipid and carbohydrate contents of tamarillo were also lower than those found in the aforementioned fruits (TACO, 2011). On the other hand, the protein, fiber, calcium, and iron contents of tamarillo were higher than those of the listed fruits (TACO, 2011).

available in the interature.	
Compounds	Tamarillo (results found in other studies)
Calories (kcal)	55.38 ³ to 64.9 ¹
Moisture (%)	82.9^1 to 88.4 ± 0.4^2
Ash (%)	0.15 ± 0.05^3 to 1.0^2
Protein (%)	1.4 ± 0.02^2 to 2.5 ± 0.19^4
Lipids (%)	0.05 ± 0.005^5 to 1.22 ± 0.29^3
Fiber (%)	0.91 ³ to 4.5 ¹
Crude Fiber	4.5 ¹
Carbohydrate (%)	3.1 ± 0.02^2 to 9.41 ± 1.76^3
Calcium (mg/ 100g)	10.6 ± 0.05^5 to 25.56^1
Iron (mg/ 100g)	0.6 ± 0.03^5 to 0.9^1
Vitamin C	16.09 ± 1.6^{1} to 19.7 ± 0.25^{5} mg/ 100g
Vitamin C by Tillmans (mg/ 100g)	16.09 ± 1.6^{1}
Total phenolic content	3.62±0.39 ⁶ g/ 100g d.m. to 130±0.08 ¹ mg GAE/100g
Antioxidant activity, DPPH	82.021 ± 7.240 mg/ g ⁷ to 853 ± 52 g equivalente trolox / g ¹
Antioxidant activity ABTS (µmol	$22 \pm 0.4 \ \mu mol \ Trolox/g \ (dm)^8$ to $853 \pm 52 \ g$ equivalente
trolox/g of sample)	trolox / g ¹

 Table 4 - Nutritional composition, phenolic compounds, and antioxidant sctivity of tamarillo available in the literature.

Studies on tamarillo: 1-De Carrasco *et al.*, 2008; 2-Diep *et al.*, 2020; 3-Pantoja *et al.*, 2009; 4-Vasco *et al.*, 2009; 5-Romero-Rodriguez *et al.*, 1994; 6-Acosta-Quezada *et al.*, 2015; 7-Ghosal *et al.*, 2013; 8-Espin *et al.*, 2016.

3.3 Nutritional composition and antioxidant activity of "prata" and "nanica" banana biomasses

A daily portion of 64 g of green banana biomass, corresponding to 70 kcal, would be recommended for nutritional provision and health promotion. From a clinical perspective, studies reported daily biomass consumption of 30 g (Cassettari *et al.*, 2019) to 300 g (Rabbani *et al.*, 2010), for benefits to gastrointestinal health (Álvarez-Acosta *et al.*, 2009; Cassettari *et al.*, 2019; Gunasekaran *et al.*, 2020; Rabbani *et al.*, 2001, 2004, 2009, 2010), body composition (Álvarez-Acosta *et al.*, 2009; Costa *et al.*, 2019; Lousek *et al.*, 2021) and lipid profile (Silva *et al.*, 2016; Lotfollahi *et al.*, 2020).

The nutritional composition of the two green banana biomasses ("prata" and "nanica") was similar, with no statistical difference (p>0.05) for: calories (110.45 and 108.11 g/ 100 kcal / 100g), ash (1.09 and 1.47 g/ 100g), and crude fiber (0.9 and 0.7 g/ 100g), respectively. (Table 5).

However, the "Nanica" banana biomass had a higher content of moisture, protein, iron, and phenolic compounds. In contrast, "Prata" banana biomass was higher in carbohydrate, calcium, vitamin C and carotenoids (p<0.05).

In this study, the values obtained for ash, protein, and fiber (Table 5) are intermediate when compared to those found in the literature (Table 6). The lipid content obtained in the biomasses of Prata and Nanica bananas were lower than those reported by all the studies in Table 6 (0.13 to 0.55, in Sena *et al.* (2020) and Silva *et al.* (2016), respectively).

The biomasses showed lower moisture values than other studies, from 71.17 to 78.96 g/100g (Bahado-Singh *et al.*, 2006 and Riquette *et al.*, 2019, respectively). In this study, during the processing of banana biomass, cooking broth was not added, which may have contributed to obtaining a product with lower moisture content. This result can favor the conservation of the product, considering that the higher moisture content can contribute to the growth of microorganisms.

The content of vitamin C found (Table 5) was lower than in Riquette *et al.* (2019), 54.4 mg/ 100g, (Table 6). The differences found can be attributed to the method for quantifying these compounds. In some studies, include in the discussion, the determination of vitamin C was carried out by HPLC. In this study, the methodology adopted was Tillman.

of frata and Namea Da	ananas.		
Compounds	"Prata" banana biomass	"Nanica"	ANOVA
	(Musa sapientum)	banana biomass	(p-value)
		(Musa Cavendish)	
Calories (kcal)	110.28±0.76	108.63±1.02	0.0875
Moisture (% w/w)	70.67±0.04	71.35±0.12	0.0007*
Ash (% w/w)	0.93±0.01	0.96±0.04	0.2500
Protein (% w/w)	1.09±0.02	1.47±0.04	0.0001*
Lipids (% w/w)	0.09±0.04	0.03±0.02	0.5900
Crude fiber (% w/w)	0.9±0.1	0.7±0.1	0.0550
Carbohydrate (% w/w)	26.28±0.09	25.32±0.31	0.0067*
Calcium (mg/ 100g)	16.33 ± 1.52	12 ± 1	0.0147*
Iron (mg/ 100g)	0.78 ± 0.02	1.06 ± 0.02	0.0001*
Vitamin C (mg/ 100g)	4.43 ± 0.31	3.72 ± 0.02	0.0166*
Carotenoids (mg/ 100g)	0.616±0.039	Nd	0.0001*
Total phenolic content (mg	51.08±6.15	99.45±7.76	0.0011*
GAE/ 100g)			
Antioxidant activity: DPPH	506±157;	1576±43;	0.0014*
(mg/g; % AA)	43.96±3.86 %	80.88±2.72 %	
Antioxidant activity: ABTS	20.95±3.3	10.03±2.55	0.0662
(mg/g)			

Table 5 - Nutritional composition, phenolic compounds, and antioxidant activity in biomasses of "Prata" and "Nanica" bananas.

Analysis on a wet basis, expressed as mean \pm standard deviation. Nd – not detected, even in more concentrated extracts. *Statistically significant difference (p < 0.05).

Table 6 - Studies on nutritional composition, j	phenolic compounds, and antioxidant activity in
green banana and green banana biomass.	

Reference	Riquette <i>et</i> <i>al.</i> (2019)	Silva et al.	Sena <i>et al.</i> (2020)	Bahado- Singh <i>et al</i> .	Bahado- Singh <i>et al</i> .	Costa <i>et</i> <i>al.</i> (2017)	Auriema <i>et</i> <i>al.</i> (2021)
	<i>al.</i> (2019)	(2016)	(2020)	(2006)	(2006)	<i>al.</i> (2017)	<i>al.</i> (2021)
Green banana	"Nanica"	"Prata"	"Terra"	Green	Green	Green	Green
	banana	banana	banana	plantain	banana	banana	banana
	biomass	biomass	biomass			pulp	biomass
Moisture (%)	78.78±0.13	77.0	75.37	68.06±0.18	71.17±0.20	76.7±0.76	78.58±0.20
Ash (%)	0.43±0.04	4.53	0.65	1.13±0.04	0.89±0.03	0.8±0.07	0.65±0.04
Protein (%)	1.44±0.13	2.73	0.81	1.67±0.02	1.42±0.18	0.8±0.04	0.94±0.04
Lipids (%)	0.20±0.01	0.505	0.13	0.24±0.02	0.33±0.03	0.2±0.07	0.40±0.03
Fibers (%)			2.55	0.48±0.03	059±0.04		4.16±0.20
Crude fiber	2.60±0.02	5.52					
(%)							
Carbohydrate (%)	16.65 ± 0.09		21.06	19.29	22.20	21.9	19.43±0.47
Calcium (mg/ 100g)							78 ± 4.90
Iron (mg/ 100g)							7.21 ± 0.29
Vitamin C (mg/100g)	54.4±0.8						
Total phenolic	322.20±1.8		189.9±15.7				
content	mg GAE/		mg				
	100 g		GAE/100g				

The carotenoid content detected in the samples was low (0.616 mg/ 100g in "Prata" banana biomass and not detected in "Nanica") and does not represent even 5% of the daily nutritional recommendations in a 100g portion (Institute of Medicine, 2019). Campos *et al.* (2024) observed low levels of carotenoids in the peel of "prata" and "nanica" green bananas and an increase during the ripening process.

The carbohydrate values in our study were higher than others, which ranged from 16,65 to 22,2 g/100g (Riquette *et al.*, 2019 and Bahado-Singh *et al.*, 2006, respectively). The carbohydrate determination calculated based on the difference in macronutrients may have overestimated the calorie content of the biomass, considering that studies indicate that green banana biomass is a source of resistant starch (Castelo-Branco *et al.*, 2017; Falcomer *et al.*, 2019), an indigestible polysaccharide, which exerts effects on the body similar to those of fiber (McCleary; Cox, 2017). This starch is physiologically analyzed as soluble fiber and chemically as insoluble fiber (Brown, 2004), which may not have been detected in our analyzes due to the crude fiber methodology (AOAC, 2005). An *in vitro* starch digestion study indicated that of the total starch present in green banana biomass, 21% was slowly digested and 42% was resistant starch (Raveena *et al.*, 2022). In a human study, green banana biomass resulted in a much lower glycemic index than ripe bananas (39 and 66, respectively), with the same cooking technique (Bahado-Singh *et al.*, 2006), highlighting the beneficial effect of resistant starch present in green banana biomass.

The variations found between the studies (Table 6) can be explained, among other factors mentioned earlier, by the use of different banana species, processing methods, and cooking times. In most studies, the fruit with the peel was cooked under pressure (Auriema *et al.*, 2021; Silva *et al.*, 2016; Riquette *et al.*, 2019; Sena *et al.*, 2020). However, Bahado-Singh *et al.* (2006) cooked peeled green bananas over low heat without pressure, and Costa *et al.* (2017) used industrialized biomass and did not provide information about the banana species or processing. The cooking times in the studies varied from 5 (Riquette *et al.*, 2019; Sena *et al.*, 2019; Sena *et al.*, 2020), 8 (Silva *et al.*, 2016), 10 (Bahado-Singh *et al.*, 2006), to 15 minutes (Auriema *et al.*, 2021).

The "Nanica" banana biomass had a higher content of antioxidant activity than in "Prata" (1576 and 506 mg/g of the sample, respectively), using the DPPH method. On the other hand, in ABTS method, it was higher in the biomass of "Prata" banana than in "Nanica" one (20.95 e 10.03 mg/g of the sample, respectively).

The antioxidant activity of cooked green "Prata" banana biomass (DPPH: 506 and ABTS: 20.95 mg/g) and the total phenolic content - TPC ("Prata": 51.08 mg GAE/ 100g) in our study were higher than that of ripe "Prata" banana (DPPH: 6.4091 and ABTS: 14.2227 mg/g; TPC: 39.69 mg GAE/ 100g) (Maduwanthi; Marapan, 2021). Campos *et al.* (2024) also observed high levels of phenolic acids in the peel of "prata" and "nanica" green bananas and a decrease during the ripening process. We did not find studies analyzing the antioxidant activity of green banana biomass of the species studied. In this way, this work contributes to reducing this knowledge gap.

3.4. Correlation between antioxidant activity and bioactive compounds

The comparative analysis of antioxidant activity and phenolic compounds faces challenges as the result is influenced by several factors, such as the extraction method (in different proportions of solvent, which can be acetone, alcohol, methanol, water), sample care (harvest, transport, storage) and the different units adopted to represent the result (Trolox equivalent per g of sample, gram of extract, % antioxidant activity, CE₅₀).

There was a strong correlation between antioxidant activity by DPPH and bioactive compounds (Phenolics: 0.89; carotenoids: 0,98). These values were higher than those observed in other studies: DPPH x total phenolic content (r = 0.7564), flavonoids (r = 0.6519), proteins (r = 0.7616) and caffeic acid (r = 0.7836) (Cruz *et al.*, 2021). Souza *et al.* (2014) also observed that the greater the content of total phenols, the greater the antioxidant activity (lower CE₅₀ value), using the DPPH method.

The correlation between antioxidant activity by DPPH and ABTS was 0.62 and non-significant (p=0.381). The variation between methods may occur due to differences in the

hydrophilic and/or lipophilic affinity of the radical cation (DPPH and ABTS) and the predominant compounds in each sample (Boroski *et al.*, 2015).

3.5. Considerations on the analyzed UFP

The four UFP had low lipid content $(0.03\pm0.02 \text{ to } 0.34\pm0.09 \text{ g}/100\text{g})$, in nanica banana biomass and ora-pro-nobis), lower than those reported in the literature (Tables 2, 4, and 6). The nanica banana biomass had the lowest lipid content $(0.03\pm0.02 \text{ g}/100\text{g})$.

Ora-pro-nobis had the highest protein content $(3.46\pm0.06 \text{ g}/100\text{g})$ among the analyzed UFP. The moisture content was similar between ora-pro-nobis and tamarillo $(87.45\pm0.07 \text{ to } 86.71\pm0.33 \text{ g}/100\text{g})$ and between the two types of biomass $(70.67\pm0.04 \text{ for prata and } 71.35\pm0.12 \text{ g}/100\text{g})$ for nanica). The ash content was similar and lower in the biomass $(1.09\pm0.02 \text{ for prata and } 1.47\pm0.04 \text{ g}/100\text{g})$ for nanica) and higher in ora-pro-nobis (2.17 ± 0.14) . Using the ABTS method, tamarillo showed the highest antioxidant activity, and "nanica" banana biomass showed the lowest $(283.81\pm3.11 \text{ and } 40.06\pm10.21 \mu \text{mol trolox/g of sample, respectively})$.

Using the DPPH method, antioxidant activity was higher in tamarillo (283.81 ± 3.11), followed by "prata" banana (83.70 ± 13.20), ora-pro-nobis (57.49 ± 3.60), and "nanica" banana (40.06 ± 10.21).

Variations in nutrient and bioactive compound levels were observed between the studies presented in the discussion. Several conditions can affect this composition, such as: the type of cultivation (natural, organic, and conventional) (Oliveira *et al.*, 2013), soil characteristics, water, external factors (Oliveira & Naozuka, 2021), exposure to sunlight during cultivation (Agostini-Costa *et al.*, 2014), among others. Processing methods during storage, freezing, lyophilization (de Souza *et al.*, 2021), analysis methods, and others can also influence the levels of compounds.

4. Conclusion

The analyzed unconventional food plants are an important source of nutrients and bioactive potential. They showed promising results regarding nutrient levels, bioactive compounds, and antioxidant activity. Tamarillo stands out as an excellent source of fiber, vitamin C, and iron, a good source of calcium, and a source of protein, in a recommended portion of 189 g of fruit. Ora-pronobis is an excellent source of calcium, iron, and carotenoids, a good source of vitamin C, and a source of protein, in a recommended portion of 79 g of fresh leaves. The "Prata" and "Nanica" banana biomasses showed similar nutrient results and stand out for their antioxidant activity, presence of phenolic compounds, and low-fat content.

The use of unconventional food plants is important for greater food diversification, preservation of plant species, rescue of traditional cuisine, and promotion of food security. The inclusion of these UFP in the diet, both for direct consumption or as an ingredient, can contribute to the provision of more sustainable, natural, and healthy foods by the food industry.

Acknowledgements

We thank Universidade Federal de Viçosa, Instituto Federal do Sudeste de Minas Gerais campus Barbacena, Coordenação de Aperfeiçoamento de Pessoal de Ensino Superior (CAPES, Brazil), Fundação de Amparo à Pesquisa do Estado de Minas Gerais (FAPEMIG), Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq). We thank João Vitor Ferreira Rivelli for his contribution in translating and revising the text

References

- Acosta-Quezada, P. G., Raigón, M. D., Riofrío-Cuenca, T., García-Martínez, M. D., Plazas, M., Burneo, J. I., ... & Prohens, J. (2015). Diversity for chemical composition in a collection of different varietal types of tree tomato (Solanum betaceum Cav.), an Andean exotic fruit. *Food chemistry*, 169, 327-335. <u>https://doi.org/10.1016/j.foodchem.2014.07.152</u>
- Agostini-Costa, T. S., Pêssoa, G. K. A., Silva, D. B., Gomes, I. S., & Silva, J. P. (2014). Carotenoid composition of berries and leaves from a Cactaceae–Pereskia sp. *Journal of functional foods*, 11, 178-184. <u>https://doi.org/10.1016/j.jff.2014.09.015</u>

- Álvarez-Acosta, T., León, C., Acosta-González, S., Parra-Soto, H., Cluet-Rodriguez, I., Rossell, M. R., & Colina-Chourio, J. A. (2009). Beneficial role of green plantain [musa paradisiaca] in the management of persistent diarrhea: A prospective randomized trial. *Journal of the American College of Nutrition*, 28(2), 169–176. https://doi.org/10.1080/07315724.2009.10719768
- AOAC (2005). Official methods of analysis (18th ed.). Gaithersburg,MD: Association of Official Analytical Chemists.
- Auriema, B. E., Braz Corrêa, F. J., Guimarães, J. de T., Soares, P. T. dos S., Rosenthal, A., Zonta, E., Rosa, R. C. C., Luchese, R. H., Esmerino, E. A., & Mathias, S. P. (2021). Green banana biomass: Physicochemical and functional properties and its potential as a fat replacer in a chicken mortadella. *LWT*, 140. <u>https://doi.org/10.1016/j.lwt.2020.110686</u>
- Bahado-Singh, P. S., Wheatley, A. O., Ahmad, M. H., Morrison, E. Y. S. A., & Asemota, H. N. (2006). Food processing methods influence the glycaemic indices of some commonly eaten West Indian carbohydrate-rich foods. *The British Journal of Nutrition*, 96(3), 476–481. <u>https://doi.org/10.1079/BJN20061792</u>
- Barreira, T. F., Paula Filho, G. X., Rodrigues, V. C. C., Andrade, F. M., Santos, R. H., Priore, S. E., & Pinheiro-Sant'ana, H. M. (2015). Diversidade e equitabilidade de plantas alimentícias não convencionais na zona rural de Viçosa, Minas Gerais, Brasil. *Revista Brasileira de Plantas Medicinais*, 17(4 suppl 2), 964-974. <u>https://doi.org/10.1590/1983-084X/14_100</u>
- Barreira, T. F., PAULA FILHO, G. X. D., Priore, S. E., Santos, R. H. S., & PINHEIRO-SANT'ANA, H. M. (2020). Nutrient content in ora-pro-nóbis (Pereskia aculeata Mill.): unconventional vegetable of the Brazilian Atlantic Forest. *Food Science and Technology*, 41(suppl 1), 47-51. <u>https://doi.org/10.1590/fst.07920</u>
- Blois, M. S. (1958). Antioxidant determinations by the use of a stable free radical. Nature, 181, 1199–1200.
- Bloor, S. J. (2001). Overview of methods for analysis and identification of flavonoids. In Methods in Enzymology. Academic Press.
- Bomfim, N. S., Ferreira, R. S., Silva e Oliveira, J., & Alfenas, R. C. G. (2024). Green banana biomass anti-obesogenic, anti-hyperlipidemic, antidiabetic, and intestinal function potential effects: a systematic review. *Nutrition Reviews*, nuae040. https://doi.org/10.1093/nutrit/nuae040
- Boroski, M., Visentainer, J. V., Cottica, S. M., & de Morais, D. R. (215 C.E.). Antioxidantes: princípios e métodos analíticos. Appris.
- Botrel, N., Freitas, S., Fonseca, M. J. D. O., Melo, R. A. D. C. E., & Madeira, N. (2020). Valor nutricional de hortaliças folhosas não convencionais cultivadas no Bioma Cerrado. *Brazilian Journal of Food Technology*, 23, e2018174. https://doi.org/10.1590/1981-6723.17418.
- Brasil. Ministério da Agricultura, P. e A. (2007). *Metodologia Científica: determinação da atividade antioxidante total em frutas pela captura do radical livre ABTS* (Vol. 128). Comunicado técnico.
- Brito, T. B. N., Pereira, A. P. A., Pastore, G. M., Moreira, R. F. A., Ferreira, M. S. L., & Fai, A. E. C. (2020). Chemical composition and physicochemical characterization for cabbage and pineapple by-products flour valorization. *LWT*, *124*, 109028. https://doi.org/10.1016/j.lwt.2020.109028
- Brown, I. L. (2004). Applications and Uses of Resistant Starch. *Journal of AOAC International*, 87(3), 727–732. <u>https://doi.org/10.1093/jaoac/87.3.727</u>
- Campos, M. T., Maia, L. F., Popović-Djordjević, J., Edwards, H. G., & de Oliveira, L. F. (2024). Ripening process in exocarps of scarlet eggplant (Solanum aethiopicum) and banana (Musa spp.) investigated by Raman spectroscopy. *Food Chemistry: Molecular Sciences*, 8, 100204. <u>https://doi.org/10.1016/j.fochms.2024.100204</u>
- Cassettari, V. M. G., Machado, N. C., Lourenção, P. L. T. de A., Carvalho, M. A., & Ortolan, E. V. P. (2019). Combinations of laxatives and green banana biomass on the treatment of functional constipation in children and adolescents: a randomized study. *Jornal de Pediatria*, 95(1), 27–33. <u>https://doi.org/10.1016/j.jped.2017.10.011</u>

- Castelo-Branco, V. N., Guimarães, J. N., Souza, L., Guedes, M. R., Silva, P. M., Ferrão, L. L., Miyahira, R. F., Guimarães, R. R., Freitas, S. M. L., Reis, M. C. dos, & Zago, L. (2017). The use of green banana (Musa balbisiana) pulp and peel flour as an ingredient for tagliatelle pasta. *Brazilian Journal of Food Technology*, 20(0). <u>https://doi.org/10.1590/1981-6723.11916</u>
- Costa, E. L., Alencar, N. M. M., Rullo, B. G. D. S., & Taralo, R. L. (2017). Effect of green banana pulp on physicochemical and sensory properties of probiotic yoghurt. *Food Science and Technology (Brazil)*, 37(3), 363–368. <u>https://doi.org/10.1590/1678-457x.01016</u>
- Costa, E. S., França, C. N., Fonseca, F. A. H., Kato, J. T., Bianco, H. T., Freitas, T. T., Fonseca, H. A. R., Figueiredo Neto, A. M., & Izar, M. C. (2019). Beneficial effects of green banana biomass consumption in patients with pre-diabetes and type 2 diabetes: A randomised controlled trial. *British Journal of Nutrition*, 121(12), 1365–1375. https://doi.org/10.1017/S0007114519000576
- Cruz, T. M., Santos, J. S., do Carmo, M. A. V., Hellström, J., Pihlava, J. M., Azevedo, L., Granato, D., & Marques, M. B. (2021). Extraction optimization of bioactive compounds from ora-pronobis (Pereskia aculeata Miller) leaves and their in vitro antioxidant and antihemolytic activities. *Food Chemistry*, 361. <u>https://doi.org/10.1016/j.foodchem.2021.130078</u>
- de Carrasco, R. R., & Zelada, C. R. E. (2008). Determinación de la capacidad antioxidante y compuestos bioactivos de frutas nativas peruanas. *Revista de la sociedad química del Perú*, 74(2), 108-124.
- De Souza, T. C. L., Da Silveira, T. F. F., Rodrigues, M. I., Ruiz, A. L. T. G., Neves, D. A., Duarte, M. C. T., ... & Godoy, H. T. (2021). A study of the bioactive potential of seven neglected and underutilized leaves consumed in Brazil. *Food chemistry*, 364, 130350. https://doi.org/10.1016/j.foodchem.2021.130350
- Diep, T. T., Pook, C., & Yoo, M. J. Y. (2020). Physicochemical properties and proximate composition of tamarillo (Solanum betaceum Cav.) fruits from New Zealand. *Journal of food composition and analysis*, 92, 103563. <u>https://doi.org/10.1016/j.jfca.2020.103563</u>
- Dutra de Oliveira, J. E., & Marchini, J. S. (1998). Ciências nutricionais. Sarvier.
- Espín, S., González-Manzano, S., Taco, V., Poveda, C., Ayuda-Durán, B., González-Paramas, A. M., & Santos-Buelga, C. (2016). Phenolic composition and antioxidant capacity of yellow and purple-red Ecuadorian cultivars of tree tomato (Solanum betaceum Cav.). *Food chemistry*, 194, 1073-1080. <u>https://doi.org/10.1016/j.foodchem.2015.07.131</u>
- Falcomer, A. L., Riquette, R. F. R., De Lima, B. R., Ginani, V. C., & Zandonadi, R. P. (2019). Health benefits of green banana consumption: A systematic review. In *Nutrients* (Vol. 11, Issue 6). MDPI AG. <u>https://doi.org/10.3390/nu11061222</u>
- Favela-González, K. M., Hernández-Almanza, A. Y., & De la Fuente-Salcido, N. M. (2020). The value of bioactive compounds of cruciferous vegetables (Brassica) as antimicrobials and antioxidants: A review. *Journal of Food Biochemistry*, 44(10), e13414. <u>https://doi.org/10.1111/jfbc.13414</u>
- Feitosa, B. F., Alcântara, C. M. de, Lucena, Y. J. A. de, Oliveira, E. N. A. de, Cavalcanti, M. T., Mariutti, L. R. B., & Lopes, M. F. (2023). Green banana biomass (Musa spp.) as a natural food additive in artisanal tomato sauce. *Food Research International*, 170. https://doi.org/10.1016/j.foodres.2023.113021
- Ferreira, R. S., Della Lucia, C. M., Minim, V. P. R., Vieira, É. N. R., Gomes, J. M. G., & Vidigal, M. C. T. R. (2024). Ora-Pro-Nobis (Pereskia aculeata) and its technological applications in foods: a review. OBSERVATÓRIO DE LA ECONOMÍA LATINOAMERICANA, 22(3), e3961e3961. <u>https://doi.org/10.55905/oelv22n3-212</u>
- Garcia, J. A., Corrêa, R. C., Barros, L., Pereira, C., Abreu, R. M., Alves, M. J., ... & Ferreira, I. C. (2019). Phytochemical profile and biological activities of Ora-pro-nobis' leaves (Pereskia aculeata Miller), an underexploited superfood from the Brazilian Atlantic Forest. *Food chemistry*, 294, 302-308. <u>https://doi.org/10.1016/j.foodchem.2019.05.074</u>
- Ghosal, M., Chhetri, P. K., Ghosh, M. K., & Mandal, P. (2013). Changes in antioxidant activity of Cyphomandra betacea (Cav.) Sendtn. fruits during maturation and senescence. *International*

Journal of Food Properties, *16*(7), 1552-1564. https://doi.org/10.1080/10942912.2011.600493

- Gunasekaran, D., Chandramohan, A., Karthikeyan, K., Balasubramaniam, B., Jagadeesan, P., & Soundararajan, P. (2020). Effect of Green Banana (Musa paradisiaca) on Recovery in Children With Acute Watery Diarrhea With No Dehydration : A Randomized Controlled Trial.
- Institute of Medicine, & National Academy of Sciences. (2019). *Dietary Reference Intakes: EAR, RDA, AI, Acceptable Macronutrient Distritubtion Ranges, and UL.* www.nap.edu
- Jacob, M. C. M., Araujo de Medeiros, M. F., & Albuquerque, U. P. (2020). Biodiverse food plants in the semiarid region of Brazil have unknown potential: A systematic review. *PLoS* One, 15(5), e0230936. <u>https://doi.org/10.1371/journal.pone.0230936</u>
- Kinupp, V. F., & Lorenzi, H. (2014). *Plantas alimentícias não convencionais (PANC) no Brasil: guia de identificação, aspectos nutricionais e receitas ilustradas*. Instituto Plantarum de estudos da flora.
- Liu, R. H. (2013). Health-promoting components of fruits and vegetables in the diet. *Advances in nutrition*, 4(3), 384S-392S. <u>https://doi.org/10.3945/an.112.003517</u>
- Lotfollahi, Z., Mello, A. P. de Q., Costa, E. S., Oliveira, C. L. P., Damasceno, N. R. T., Izar, M. C., & Neto, A. M. F. (2020). Green-banana biomass consumption by diabetic patients improves plasma low-density lipoprotein particle functionality. *Scientific Reports*, 10(1). https://doi.org/10.1038/s41598-020-69288-1
- Lousek, N. F., Santos, N. C., Dourado, M. C. M., Pontieri, F. M., Teófilo, M. N. G., Castro, F. S., Costa, S. H. N., Blanch, G. T., Borges, A. F., & Gomes, C. M. (2021). Effects of Green Banana Biomass (Musa spp.) on Laboratory Parameters of Animal Models of Wistar Mice under Hyperlipidic Diet. *Journal of the American College of Nutrition*, 40(5), 472–477. <u>https://doi.org/10.1080/07315724.2020.1792811</u>
- Maciel, V. B. V., Bezerra, R. Q., Chagas, E. G. L. D., Yoshida, C. M. P., & Carvalho, R. A. D. (2021). Ora-pro-nobis (Pereskia aculeata Miller): a potential alternative for iron supplementation and phytochemical compounds. *Brazilian Journal of Food Technology*, 24, e2020180. <u>https://doi.org/10.1590/1981-6723.18020</u>
- Maduwanthi, S. D. T., & Marapana, R. A. U. J. (2021). Total phenolics, flavonoids and antioxidant activity following simulated gastro-intestinal digestion and dialysis of banana (Musa acuminata, AAB) as affected by induced ripening agents. Food Chemistry, 339, 127909. <u>https://doi.org/10.1016/j.foodchem.2020.127909</u>
- MAPA. Ministério da Agricultura, Pecuária e Abastecimento (2010). *Manual de hortaliças não convencionais*. MAPA, Brasília.
- Martin, D., Lopes, T., Correia, S., Canhoto, J., Marques, M. P. M., & de Carvalho, L. A. B. (2021). Nutraceutical properties of tamarillo fruits: A vibrational study. *Spectrochimica Acta Part A: Molecular* and *Biomolecular* Spectroscopy, 252, 119501. https://doi.org/10.1016/j.saa.2021.119501
- McCleary, B. V, & Cox, J. (2017). Evolution of a Definition for Dietary Fiber and Methodology to Service this Definition. *Luminacoids Research*, 26(2), 9–21.O'Callaghan, D.J. & Guinee, T.P. (2004) Rheology and texture of cheese. In: Fox, P.F., McSweeney, P.L.H., Cogan, T.M.C., Guinee, T.P. (Eds.) *Cheese: Chemistry, Physics and Microbiology* (3rd), Academic Press, New York, NY, 511-540.
- Monteiro, R. L., Garcia, A. H., Tribuzi, G., Mattar Carciofi, B. A., & Laurindo, J. B. (2021). Microwave vacuum drying of Pereskia aculeata Miller leaves: Powder production and characterization. *Journal of Food Process Engineering*, 44(2), e13612. <u>https://doi.org/10.1111/jfpe.13612</u>
- Oliveira, D. D. C. D. S., Wobeto, C., Zanuzo, M. R., & Severgnini, C. (2013). Composição mineral e teor de ácido ascórbico nas folhas de quatro espécies olerícolas nãoconvencionais. *Horticultura Brasileira*, 31, 472-475. <u>https://doi.org/10.1590/S0102-05362013000300021</u>

- Oliveira, H. A. B., Anunciação, P. C., Silva, B. P., Souza, Â. M. N., Pinheiro, S. S., Lucia, C. M. D., Cardoso, L. M., Castro, L. C. V., & Pinheiro-Sant'Ana, H. M. (2019). Nutritional value of non-conventional vegetables prepared by family farmers in rural communities. *Ciência Rural*, 49(8), e20180918–e20180918. https://doi.org/10.1590/0103-8478cr20180918
- Oliveira, A. P. D., & Naozuka, J. (2021). Iron species and proteins distribution in unconventional food plants. Brazilian Journal of Food Technology, 24, e2020294. https://doi.org/10.1590/1981-6723.29420
- Orqueda, M. E., Torres, S., Verón, H., Pérez, J., Rodriguez, F., Zampini, C., & Isla, M. I. (2021). Physicochemical, microbiological, functional and sensory properties of frozen pulp of orange and orange-red chilto (Solanum betaceum Cav.) fruits. Scientia Horticulturae, 276, 109736. <u>https://doi.org/10.1016/j.scienta.2020.109736</u>
- Pantoja, L., Pinto, N. A. V. D., Lopes, C., Gandra, R., & Santos, A. S. D. (2009). Caracterização física e físico-química de frutos de duas variedades de tamarilho oriundas do norte de Minas Gerais. *Revista Brasileira de Fruticultura*, 31, 916-919. <u>https://doi.org/10.1590/S0100-29452009000300041</u>
- Philippi, S. T. (2008). Pirâmide dos alimentos: fundamentos básicos da nutrição. Manole, Barueri.
- Sant'Ana, H. M. P., Stringheta, P. C., Brandão, S. C. C., & de Azeredo, R. M. C. (1998). Carotenoid retention and vitamin A value in carrot (Daucus carota L.) prepared by food service. Food Chemistry, 61(1-2), 145-151. <u>https://doi.org/10.1016/S0308-8146(97)00084-8</u>
- Porto, F. G. S., Campos, Â. D., Carreño, N. L. V., & Garcia, I. T. S. (2021). Pereskia aculeata leaves: properties and potentialities for the development of new products. Natural Product Research, 36(18), 4821-4832. <u>https://doi.org/10.1080/14786419.2021.2010070</u>
- Rabbani, G. H., Ahmed, S., Hossain, M. I., Islam, R., Marni, F., Akhtar, M., & Majid, N. (2009). Green banana reduces clinical severity of childhood shigellosis: A double-blind, randomized, controlled clinical trial. *Pediatric Infectious Disease Journal*, 28(5), 420–425. <u>https://doi.org/10.1097/INF.0b013e31819510b5</u>
- Rabbani, G. H., Larson, C. P., Islam, R., Saha, U. R., & Kabir, A. (2010). Green bananasupplemented diet in the home management of acute and prolonged diarrhoea in children: A community-based trial in rural Bangladesh. *Tropical Medicine and International Health*, 15(10), 1132–1139. <u>https://doi.org/10.1111/j.1365-3156.2010.02608.x</u>
- Rabbani, G. H., Teka, T., Kumar Saha, S., Zaman, B., Majid, N., Khatun, M., Wahed, M. A., & Fuchs, G. J. (2004). Green Banana and Pectin Improve Small Intestinal Permeability and Reduce Fluid Loss in Bangladeshi Children with Persistent Diarrhea. In *Digestive Diseases* and Sciences (Vol. 49, Issue 3).
- Rabbani, G. H., Teka, T., Zaman, B., Majid, N., Khatun, M., & Fuchs, G. J. (2001). Clinical studies in persistent diarrhea: Dietary management with green banana or pectin in Bangladeshi children. *Gastroenterology*, 121(3), 554–560. <u>https://doi.org/10.1053/gast.2001.27178</u>
- Raveena, N. K., Ingaladal, N., Reshma, M. V., & Lankalapalli, R. S. (2022). Phytochemical investigation of unripe banana (Musa AAB) cv. Nendran and its novel 'Banana Grits.' *Food Chemistry Advances*, 1. <u>https://doi.org/10.1016/j.focha.2022.100063</u>
- Riquette, R. F. R., Ginani, V. C., Leandro, E. dos S., de Alencar, E. R., Maldonade, I. R., de Aguiar, L. A., de Souza Acácio, G. M., Mariano, D. R. H., & Zandonadi, R. P. (2019). Do production and storage affect the quality of green banana biomass? *LWT*, 111, 190–203. <u>https://doi.org/10.1016/j.lwt.2019.04.094</u>
- Rodriguez-Amaya, D. B. (2001). A guide to carotenoid analysis in foods. Ilsi Press Washington Ed.
- Romero-Rodriguez, M. A., Vazquez-Oderiz, M. L., Lopez-Hernandez, J., & Simal-Lozano, J. (1994). Composition of babaco, feijoa, passion-fruit and tamarillo produced in Galicia (NW Spain). Food Chemistry, 49, 3, 251-255.
- Santos, P. P. A. D., Ferrari, G. D. S., Rosa, M. D. S., Almeida, K., Araújo, L. D. A. D., Pereira, M. H. C., ... & Morato, P. N. (2022). Desenvolvimento e caracterização de sorvete funcional de alto teor proteico com ora-pro-nóbis (Pereskia aculeata Miller) e inulina. Brazilian Journal of Food Technology, 25, e2020129. <u>https://doi.org/10.1590/1981-6723.12920</u>

- Sarruge, J. R., & Haag, H. P. (1974). *Análise química de plantas*. Escola Superior de Agricultura Luiz de Queiroz.
- Sena, L. de O., Viana, E. D. S., Reis, R. C., Barreto, N. S. E., Sampaio De Santana, T., & Assis, J. L. de J. (2020). COMUNICADO TÉCNICO 171 (Comunicado técnico). Embrapa.
- Singleton, V. L., Orthofer, R., & Lamuela-Raventós, R. M. (1999). Analysis of total phenols and other oxidation substrates and antioxidants by means of folin-ciocalteu reagent. *Methods in Enzymology*, 299, 152–178.
- Silva, A. R., Cerdeira, C. D., Brito, A. R., Salles, B. C. C., Ravazi, G. F., Moraes, G. de O. I., Rufino, L. R. A., de Oliveira, R. B. S., & Santos, G. B. (2016). Green banana pasta diet prevents oxidative damage in liver and kidney and improves biochemical parameters in type 1 diabetic rats. *Archives of Endocrinology and Metabolism*, 60(4), 355–366. https://doi.org/10.1590/2359-3997000000152
- Silveira, M. G., Picinin, C. T., Cirillo, M. A., Freire, J. M., & Barcelos, M. D. F. P. (2020). Nutritional assay Pereskia spp.: unconventional vegetable. *Anais da Academia Brasileira de Ciências*, 92(suppl 1), e20180757. <u>https://doi.org/10.1590/0001-3765202020180757</u>
- Souza, L. F., De Barros, I. B. I., Mancini, E., De Martino, L., Scandolera, E., & De Feo, V. (2014). Chemical composition and biological activities of the essential oils from two pereskia species grown in Brazil. *Natural Product Communications*, 9(12), 1805–1808. <u>https://doi.org/10.1177/1934578x1400901237</u>
- TACO (2011) Tabela brasileira de composição de alimentos (4th ed.). NEPA-UNICAMP, Campinas. Retrieved from https://www.gov.br/agricultura/pt-br/assuntos/inspecao/produtosvegetal/legislacao-de-produtos-origem-vegetal/biblioteca-de-normas-vinhos-ebebidas/tabela-brasileira-de-composicao-de-alimentos_taco_2011.pdf Accessed Ago 19, 2024.
- Takeiti, C. Y., Antonio, G. C., Motta, E. M., Collares-Queiroz, F. P., & Park, K. J. (2009). Nutritive evaluation of a non-conventional leafy vegetable (Pereskia aculeata Miller). *International Journal of Food Sciences and Nutrition*, 60(sup1), 148-160. https://doi.org/10.1080/09637480802534509
- Vasco, C., Avila, J., Ruales, J., Svanberg, U., & Kamal-Eldin, A. (2009). Physical and chemical characteristics of golden-yellow and purple-red varieties of tamarillo fruit (Solanum betaceum Cav.). *International Journal of Food Sciences and Nutrition*, 60(sup7), 278-288. <u>https://doi.org/10.1080/09637480903099618</u>
- Zem, L. M., Helm, C. V., Henriques, G. S., Cabrini, D. D. A., & Zuffellato-Ribas, K. C. (2017). Pereskia aculeata: biological analysis on wistar rats. *Food Science and Technology*, 37, 42-47. <u>https://doi.org/10.1590/1678-457x.29816</u>